AMENDMENTS

IN THE SPECIFICATION

Please replace the paragraph beginning on page 1, line 5, with the following rewritten paragraph:

This application is a continuation of application Serial No. 09/829,507, filed April 9, 2001 which is a continuation of application Serial No. 09/220,265, filed December 22, 1998, which application is a continuation in part application of serial no. 08/740,947, filed November 5, 1996 (now U.S. Patent No. 5,834,593 issued November 10, 1998) all of which are incorporated herein by reference in their entirety and to which applications we claim priority under 35 U.S.C. §120.

Please replace the paragraph beginning on page 1, line 10, with the following rewritten paragraph:

The United States Government may have certain rights in this application pursuant to Grant No. AG10770 awarded by the National Institutes of Health.

Please replace the line on page 22, line 14, with the following:

ATAACTTCGTATA ATGTATGC TATACGAAGTTAT (SEQ ID NO:1)

Please replace the line on page 22, line 20, with the following:

ACAACTTCGTATA ATGTATGC TATACGAAGTTAT (SEQ ID NO:2)

Please replace the line on page 23, line 3, with the following

ATAACTTCGTATA ATGTATAC TATACGAAGTTAT (SEQ ID NO:3)

Please replace the paragraph beginning on page 23, line 8, with the following rewritten paragraph:

The lox target is placed into the genome of cultured HeLa cells using methods known in the art. The lox-neo target is placed into the HeLa cells by electroporating 10^7 cells in 0.8 ml with 1 μg of a site specific integration vector, pSF1, containing lox P sequences and an

unmodified neo gene sequence (Fukushige (1992) *PNAS* 89: 7905-7909). Electroporation uses a single pulse of 450 V at 500 μF from a BioRad Gene Pulser™. One day later, cells are selected for growth in an α^- medium which lacks deoxyribonucleosides and ribonucleosides supplemented with 15% dialyzed fetal bovine serum.

Please replace the paragraph beginning on page 27 at line 17 and continuing to the end of page 27 at line 25 with the following rewritten paragraph:

The light and heavy chain expression vectors are co-transfected into the COS-PrP knock-out line by electroporation. Equal amounts of each plasmid DNA (10 μg) are added to 0.8 ml of cells suspended in PBS at 1 X 10^7 ml^{-1}. A pulse is delivered at 1.9kV, 25 μF capacitance using a Gene Pulsar™ apparatus (BioRad). After a 10 minute recovery period at room temperature, the electroporated cells are added to 20 ml DMEM containing 10% gamma globulin-free fetal calf serum (GIBCO™). After a 2 hour incubation, the medium is collected, centrifuged to remove cellular debris, and applied to a Protein A agarose column (Affi-Gel Protein A MAPSII™ kit, BioRad) equilibrated with binding buffer. The elute is concentrated, and the buffer changed to PBS using a microconcentrator.

Please replace the paragraph beginning on page 29, lines 13-19 with the following rewritten paragraph:

The light and heavy chain expression vectors are then co-transfected into the COS-PrP tet-inducible knock-out line by electroporation. Equal amounts of each plasmid DNA (10 μg) are added to 0.8 ml of cells suspended in PBS at 1 X 10^7 ml^{-1}. A pulse is delivered at 1.9kV, 25 μF capacitance using a Gene Pulsar™ apparatus (BioRad). After a 10 minute recovery period at room temperature, the electroporated cells are added to 20 ml DMEM containing 10% gamma globulin-free fetal calf serum (GIBCO™). After a 2 hour incubation, the medium is collected, centrifuged to remove cellular debris, and purified.

Please insert the attached "Sequence Listing" as separately numbered page 1 after the abstract.